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**Please find below and/or attached an Office communication concerning this application or proceeding.**

The time period for reply, if any, is set in the attached communication.

## Office Action Summary

Application No.

10/611,440

Applicant(s)

BERINSTEIN ET AL.

Examiner

Karen A. Canella

Art Unit

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-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

### Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

### Status

- 1) ☐ Responsive to communication(s) filed on \_\_\_\_.
- 2a) ☐ This action is **FINAL**. 2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

### Disposition of Claims

- 4) ☐ Claim(s) 1-66 is/are pending in the application.
- 4a) Of the above claim(s) 36, 38 and 39 is/are withdrawn from consideration.
- 5) ☐ Claim(s) \_\_\_\_ is/are allowed.
- 6) ☐ Claim(s) 1-35, 37 and 40-66 is/are rejected.
- 7) ☐ Claim(s) \_\_\_\_ is/are objected to.
- 8) ☐ Claim(s) \_\_\_\_ are subject to restriction and/or election requirement.

### Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on \_\_\_\_ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.  
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).  
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

### Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some \* c) ☐ None of:
- ☐ Certified copies of the priority documents have been received.
  - ☐ Certified copies of the priority documents have been received in Application No. \_\_\_\_.
  - ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

\* See the attached detailed Office action for a list of the certified copies not received.

### Attachment(s)

- 1) ☒ Notice of References Cited (PTO-892)
- 2) ☐ Notice of Draftsperson's Patent Drawing Review (PTO-948)
- 3) ☒ Information Disclosure Statement(s) (PTO/SB/08)  
Paper No(s)/Mail Date Aug. 16, 2006.
- 4) ☐ Interview Summary (PTO-413)  
Paper No(s)/Mail Date. \_\_\_\_.
- 5) ☐ Notice of Informal Patent Application
- 6) ☐ Other: \_\_\_\_.

### **DETAILED ACTION**

Acknowledgment is made of applicant's election with traverse of Group I and the species of avipox. The traversal is on the grounds that it would not be undue experimentation to search for all of the groups and species together. This has been considered and found partially persuasive. Groups III, IV and VI will be rejoined to group I in order to advance prosecution. Groups drawn to SEQ ID NO:3 will not be rejoined because a search for SEQ ID NO:3 is not commensurate with a search for SEQ ID NO:1. It is noted that a search for SEQ ID NO:1 did not result in a hit on the instant SEQ ID NO:3. Thus the nucleic acids of SEQ ID NO:1 and 3 encode structurally different unrelated proteins. The peptides of BFA4 and BCY1 (groups V and VII) will not be rejoined because the search for a method of immunizing a host against the tumor antigen BFA4 comprising administering immunogenic peptide from BFA4 is not commensurate with a search for peptides derived from BFA4 or BCY1. Further the peptides of Groups V and VII could have completely different uses from the method of immunizing a host against BFA4 and different questions of patentability would have to be considered. Clearly there would be undue burden to search all of the inventions as relating to nucleic acids of SEQ ID NO:3, peptides derived from both BFA4 and BCY1 and a method of immunizing a host against BCY1.

Applicant argues that it would not be undue burden to examine all of the species of expression vector. This has been considered and found persuasive. The species election requirement is withdrawn.

The restriction requirement between the instant rejoined claims and the inventions of Groups II, V, VII and VIII is deemed proper and adhered to. The restriction requirement is hereby made FINAL.

Claims 1, 6, 20-26 have been amended. Claims 40-66 have been added. Claims 1-66 are pending. Claims 36, 38 and 39, drawn to non-elected inventions, are withdrawn from consideration. Claims 1-35, 37 and 40-66 are examined on the merits.

### ***Claim Rejections - 35 USC § 112***

The following is a quotation of the second paragraph of 35 U.S.C. 112:

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The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

Claims 21-25, 31-35 and 59-63 are rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claim 31 is vague and indefinite in the recitation of “nucleic acid sequence illustrated in SEQ ID NO:25 or 27” because SEQ UD NO:25 and 27 are peptides. For purpose of examination, the claim will be read as nucleic acid sequence encoding SEQ ID NO:25 or 27.

Claims 21 and 59 are vague and indefinite in the recitation of “co-stimulatory component” without a specific reference as to what is being stimulated

The following is a quotation of the first paragraph of 35 U.S.C. 112:

The specification shall contain a written description of the invention, and of the manner and process of making and using it, in such full, clear, concise, and exact terms as to enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the same and shall set forth the best mode contemplated by the inventor of carrying out his invention.

Claims 4-35, 37, 40-43, 47-66 are rejected under 35 U.S.C. 112, first paragraph, as failing to comply with the enablement requirement. The claim(s) contains subject matter which was not described in the specification in such a way as to enable one skilled in the art to which it pertains, or with which it is most nearly connected, to make and/or use the invention.

The factors considered when determining if the disclosure satisfies the enablement requirement and whether any necessary experimentation is undue include, but are not limited to: 1) nature of the invention, 2) state of the prior art, 3) relative skill of those in the art, 4) level of predictability in the art, 5) existence of working examples, 6) breadth of claims, 7) amount of direction or guidance by the inventor, and 8) quantity of experimentation needed to make or use the invention. In re wands, 858 F.2d 731, 737.8 USPQ2d 1400, 1404 (Fed. Cir. 1988).

(A)As drawn to a method of treating cancer comprising providing tumor antigens by administering peptides or by administering expression vectors encoding peptides

Claims 6-25, 40-43, 49-63 are drawn to the expression vector of claim 1 further comprising at least one nucleic acid encoding an additional tumor antigen and/or angiogenesis-associated antigen and/or co-stimulatory component. Claims 26-30 and 64-66 are drawn to pharmaceutical composition comprising nucleic acid expression vectors. Claims 31-35 are drawn in part to a method of treating cancer comprising administering a expression vector encoding SEQ ID NO:25 or 27. Claim 37 is drawn to a method for immunizing a host against the tumor antigen BFA4 comprising administering a peptide shown in Table V, VI or VII.

The specification teaches that six clones were used in a BFA4 peptide-pulsed target experiment (page 35-36, bridging sentence). The specification provides 100 nonamer peptides selected for their "potential ability" to bind to HLA-A\*0201 (Table V, pages 37-38). The specification states that pooled groups of the selected peptides were used to activate CTL which lysed target cells bearing the peptides (page 41, first paragraph). The specification teaches that single peptides from each group were tested to reveal a number of individual strongly reactive peptides recognized by human T cells and able to induce CTL activity in vitro (page 41, second paragraph). The specification is not enabling for administering the expression vectors to treat cancer by inducing a therapeutic immune response for the reasons set forth below nor is it enabling for a method of treating cancer comprising the administration of peptides derived from BFA4. Claims drawn to an expression vector which further incorporates tumor antigens, angiogenesis associated antigens or co-stimulatory molecules are included with the rejection as well as pharmaceutical composition because said products are clearly intended to be used for generating a therapeutic immune response to a BFA4 expressing cancer and the specification does not teach how to use such an expression vector, encoding molecules important for the immune response .

(B)As drawn to the administration of expression vectors and pharmaceutical compositions comprising expression vectors.

Claims 31-35 are drawn to the administration of expression vectors for a therapeutic effect against cancers. 6-30, 40-43 and 49-66 are drawn to expression vectors in pharmaceutical compositions and expression vectors encoding BFA4 in addition to other tumor antigens, angiogenesis associated antigens or co-stimulatory molecules all of which are intended to increase the therapeutic anti-cancer effect of an immune response against the BFA4 antigen in

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vivo. Claims 4, 5, 9, 10, 14, 15, 19, 20, 24, 25, 29, 30, 34, 35, 47, 48, 52, 53, 57, 58, 62 and 63 are included with this rejection because avian viral vectors would not be expected to be used as a recombinant vectors for the simple production of protein in vitro because said avian vectors are not capable of continued infection in mammalian cells (Tartaglia et al, Vaccine, 2001, Vol. 19, pp. 2571-2575) and thus would only be used for in vivo experimentation in mammals.

(C)As drawn to the administration of expression vectors to a patient

Claims 31-35 are drawn to the administration of an expression vector encoding SEQ ID NO:25 or 27 to a patient. Claims 6-30, 40-43, 49-66 are drawn to expression vectors and pharmaceutical compositions clearly intended for in vivo use as for the induction of an anti-tumor immune response.

The instant specification does not teach how to overcome problems with in vivo delivery and expression with respect to the administration of the claimed expression vectors or viral vectors comprising the nucleic acids encoding BFA4. . The state of the art is that in vivo gene delivery is not well developed and is highly unpredictable. For instance Verma et al (Nature, 1997, Vol. 389, pp. 239-242) teach that the Achilles heel of gene therapy is gene delivery. The specification is not enabling for the selective targeting of an antigen-presenting cells effective to mount an immune response against a generic "cancer". and without teaching regarding how said construct is to be specifically targeted one of skill in the art would be subject to undue experimentation. Verma et al further state that an ongoing problem is the inability to deliver genes efficiently and to obtain sustained expression (page 239, column 3). The specification does not teach a dosage of plasmid or viral vector which could be used as an "efficient dose" in order to inhibit the solid tumors, nor does the specification disclose a method to obtain sustained expression of the BFA4 antigen. Eck et al (Gene-Based Therapy, In: The Pharmacological Basis of Therapeutics, Goodman and Gilman, Ed.s, 1996, pp. 77-101) teach that the fate of the DNA vector itself with regard to the volume of distribution, rate of clearance into tissues etc., the in vivo consequences of altered gene expression, the fraction of vector taken up by the target cell population, the trafficking of the genetic material within cellular organelles, the rate of degradation of the DNA the level of mRNA produced, the stability of the mRNA produced in vivo are primary considerations regarding effective therapy. Eck et al state that these factors differ dramatically on the vector used, the sequence being expressed, and the disease being

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treated (Eck et al bridging pages 81-82). Thus, one of skill in the art would conclude that there is no nexus between the transfection of cell in vitro with constructs encoding BFA4, and the successful modulation of the immune response against the BFA4 antigen sufficient to cause a therapeutic effect.

Using viral vectors to deliver anti-sense DNA to an organism in vivo is in the realm of gene therapy, and therefore highly unpredictable in view of the complexity of in vivo systems. Orkin states ( "Report and Recommendation of the Panel to Assess the NIH Investment in Research on Gene Therapy", NIH, 1995) that clinical efficacy had not been definitively demonstrated with any gene therapy protocol (page 1, second paragraph). Orkin concludes that, "none of the available vector systems is entirely satisfactory, and many of the perceived advantages of vector systems have not been experimentally validated. Until progress is made in these areas, slow and erratic success in applying gene transfer methods to patients can be expected".. Orkin teaches that adequate expression of the transferred genes is essential for therapy, but that data regarding the level and consistency of expression of transferred genes in animal models was unknown. Orkin states that in protocols not involving ex vivo infections/transfection, it is necessary to target the expression of the transferred genes to the appropriate tissue or cell type by means of regulatory sequences in gene transfer vectors. As stated above, the specification does not teach a vector having a specific regulatory sequence which would direct the expression of the BFA4 nucleic acids to the appropriate tissue and/or cell type.

It is noted that Ghose et al (Human Gene Therapy, 2000, Vol. 11, pp. 1289-1301) teach that mice receiving tumor cells infected with ALVAC viral vectors did not develop tumors versus control mice which were injected with tumor cells alone. However, it is well recognized in the art that clinical results on patients do not reflect the results of animal models. For example Schultze et al (Trends in Immunology, 2004, Vol. 25, pp 659-664) teach that encouraging animal model studies lead to clinical trials, but that the general outcomes of these trials are disappointing, citing a discrepancy between the outcome of pre-clinical models and the outcome of the human situation. Bodey et al, (Anticancer Research, 2000, Vol. 20, pp. 2665-2676) teach that the animal models often produce highly encouraging results but that the resulting response in humans is disappointing. Le Fur et al (PNAS, 1997, Vol. 94, pp. 7561-7565) teach that results

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pertaining to the rejection of transplanted tissue differs from raising an immune response in a patient against a primary tumor in its natural place (page 7564, second column, lines 13-15 in the third full paragraph). Le Fur et al conclude that many practical issues need to be resolved before an effective peptide-antigen tumor vaccine is obtained from peptides identified by T cell recognition or predicted by over expressed RNA isoforms in tumors (page 7565 last paragraph). Donnelly et al (Journal of Immunology, 2005 Jul 15, 175(2):633-639 address the lack of nexus between the outcome of DNA vaccines in small mammals and humans, concluding (three years after the filing date of the instant application) that there is a disappointing potency of DNA vaccines in humans. This is corroborated by Vanniasinkam et al (Journal of Clinical Virology, 2006 Aug, 36(4):292-297) who state that ability to induce an effective immune response as a result of DNA vaccination in large animals and humans is disappointing.

The specification does not remedy any of the deficiencies or the prior art with regard to the administration of nucleic acids in vivo. Given the lack of any guidance from the specification on any of the above evidence of unreliability on the art, one of skill in the art would be subject to undue experimentation without reasonable expectation of success in order to practice the claimed methods.

(C)As drawn to the administration of tumor antigen peptides

The prior art teaches that tumor cells are phenotypically less stable than normal cells and can escape the immune response of the host by many mechanisms including deficient antigen processing by tumor cells, production of inhibitory substances such as cytokines, tolerance induction, rapidly growing cells which can overwhelm a slower immune response, failure of the host to respond to an antigen due to immunosuppression, tumor burden, infections or age, deficient antigen presentation with the host and failure of the host effector cells to reach the tumor due to the stromal barrier (Paul, Fundamental Immunology, (text), 1993, page 1163, second column, first sentence under the heading "Factors Limiting Effective Tumor Immunity" and Table 4 and Finke et al, Immunology Today, 1999, Vol. 20, pp. 158-160, see page 159, under the heading "Barriers that prevent tumor recognition"). The specification has stated that certain of the disclosed peptides are able to activate CTL which lyse BFA4 expressing cells in vitro. Paul teaches that lymphocytes from tumor bearing patients have frequently been found to be cytotoxic to their own tumor cells in vitro, but that this effect was blocked by the addition of



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sera from said patients. Paul teaches that the constituent of the sera which caused the blocking of the cytotoxicity was unknown, but that antibodies, antibody-antigen complexes and shed antigen have all been implicated in the blocking phenomenon (Paul page 1167, second paragraph under the heading "Immunological Enhancement and Blocking Factors". Finke et al (ibid) teach that tumor derived factors can alter T-cell function rendering tumor infiltrating lymphocytes dysfunctional (page 159, third column) and that an additional escape mechanism is stimulation of the Fas ligand by the tumor cells (page 159, third column). Finke et al teach that removal of the dysfunctional infiltrating lymphocytes from the tumor results in restoration of T-cell function directly implicating tumor, stoma or other host infiltrating cells on the repression of T-cell function (page 160 first column, lines 4-12). Paul also notes that in some cases, immune response to a tumor antigen may sometimes stimulate the growth of the tumor cells directly (last line under the heading "Immunological Enhancement and Blocking Factors", page 1167). With respect to the blocking factor found in serum, Apostolopoulos et al (Nature Medicine, 1998, vol. 4, pp. 315-320) teach that endogenous antibodies present at the time of administration of a tumor peptide re-routes the immune response from a cellular response to a humoral response. In preclinical experiments with mice, MUC1 peptides targeted to the mannose receptor produce high levels of CTL and a low level of antibodies. However, in human clinical trials a low level of CTL and a high level of humoral response was observed (Apostolopoulos, page 315, first column, bridging paragraph). Apostolopoulos et al teach that the presence of endogenous antibodies which bind to the MUC1 peptide was responsible for this re-routing of the immune response from cellular to humoral due to the Fc portion of the antibody (page 319, first column, lines 7-10). Apostolopoulos et al teach that mice are devoid of these antibodies (page 315, second column, lines 9-13) and are thus able to effectively mount a cellular immune response against the target antigen. Apostolopoulos et al teach that these findings have implication for other immunotherapy approaches (page 318, lines 4-8, under the heading "Discussion". In support of these conclusions Jager et al (PNAS, 2000, Vol. 97, pp. 12198-12203) teach that patients who do not have antibodies to the cancer testis antigen, NY-ESO-1, were able to generate a specific T-cell response to NY-ESO after intradermal administration, whereas patients having antibodies prior to treatment which reacted with said antigen already had T-cells which reacted with target cells expressing said antigen in vitro, and said positive patients did not

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develop significant CTL in response to the administered NY-ESO antigen. Finke et al (ibid) conclude that the immune dysfunction resulting from a progressively growing tumor is distinct from generalized immune suppression induced by pharmacological means and as such is not understood (page 160 under the heading of "Conclusion"). These references serve to demonstrate that the induction of a anti-tumor CTL response after the administration of a tumor peptide is unpredictable.

Paul (ibid) states that deficient antigen presentation is a mechanism by which tumor cells escape immune detection. This is corroborated by the observations set forth in the abstract of Semino et al (Journal of Biological Regulators and Homeostatic Agents, 1993, Vol. 7, pp. 99-105) and the abstract of Algarra et al International Journal of Clinical and Laboratory Research, 1997, Vol. 27, pp. 95-102) which all teach that primary tumors in situ are often heterogeneous with respect to MHC presentation. The effect of the claimed vaccine upon such a heterogeneous tumor has not been demonstrated by the specification. More currently, the abstract of Bodey et al (Anticancer Research, 2000 Jul-Aug, Vol. 20, pp. 2665-2676) teaches that the failure of methods of treating cancer comprising the administration of tumor antigens is due to the failure of cancer vaccines to eliminate the most dangerous cells within a tumor which are so de-differentiated that they no longer express cancer cell specific molecules.

Paul (ibid) states that the induction of tolerance is a mechanism by which tumor cells escape immune detection. The art recognizes that T-cell are subject to clonal deletion within the thymus of a host and that this mechanism eliminates T-cell which are reactive with self-antigens. Lauritzsen et al (International Journal of Cancer, 1998, Vol. 78, pp. 216-222) teach that clonal deletions of thymocytes is a major event in T-cell tolerance which could lead to a tumor escape mechanism. In transgenic mice homozygous for HLA-specific CD+4 T-cells which are specific for a MOPC315 plasmacytoma, injection of a large number of tumor cells results in apoptosis of immature and mature transgenic CD+4+8 and CD+4 thymocytes. This negative selection was specific for the transgenic thymocytes that would complement the idotype of the immunoglobulins of the MOPC315 plasmacytoma, because injection of tumor cells from a plasmacytoma which had a different idotype of immunoglobulins failed to elicit the clonal deletion. Lauritzsen et al teach that injection of purified MOPC315 protein, versus the tumor cells, caused a profound reduction of the specific thymocytes specific to the idotype of the

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plasmacytoma. Lauritzsen et al conclude that deletion of tumor specific thymocytes may represent a major escape mechanism in patients with cancers that secrete or shed antigens. In the instant case, the antigens are known self antigens. It would be reasonable to conclude that said normal antigens are presented within the thymus to developing thymocytes and T-cells with high affinity for said antigens are deleted as "self". It would be also reasonable to conclude that administration of the claimed polypeptides or cells expressing said polypeptides would not result in an efficacious vaccine as a T-cell response would not be evoked due to the process of clonal deletion in the thymus, rendering the host devoid of T-cells which are specific to the self-protein. Sarma et al (Journal of Experimental Medicine, 1999, Vol. 189, pp. 811-820) states that a critical issue in therapeutic regimens comprising the administration of tumor antigens for immunotherapy is whether un-mutated tumor antigens which are expressed in normal cells impose special restrictions on the CTL response in vivo. Using transgenic mice wherein the antigen specific T cells specific for the P1A non-mutated tumor antigen are expressed at high levels and remain responsive to the P1A antigen when assayed in vitro, it was found that P1A antigen expressed in the thymus resulted in clonal deletion of said specific T-cells. Sarma et al note that although said transgenic mice produce an overwhelming majority of T cells that are specific for P1A, said mice are no more resistant to cells expressing P1A than non-transgenic litter mates. Sarma et al concludes that even though P1A can be a tumor rejection antigen, the effector function of P1A specific CTL is restrained in vivo and that these results have important implications for the strategy of tumor immunotherapy. Further, the presence of CTL which can lyse target cells in vitro has no apparent nexus with anti-tumor cytolytic activity in vivo. Ohlen et al (Journal of Immunology, 2001, Vol. 166, pp. 2863-2870) teach that T-cells recognizing normal proteins expressed in tumors can be isolated in vitro, but that the existence of said T-cells does not preclude in vivo anergy induction and deletion (page 2863, second column, lines 1-6 of the last paragraph). Antoinia et al (International Immunology, 1995, Vol. 7, pp. 715-725) teach that T-cells which are impaired in the ability to proliferate in response to antigen and unable to reject tumors in vivo were fully functional as CTL lymphocytes in vivo (page 724, first column, first full paragraph). Yu and Restifo (Journal of Clinical Investigation, 2002, Vol. 110, pp. 289-294, especially page 292) teach that even when increased anti-tumor T-cell precursors have been induced by vaccination, the clinical response is partial and transient and most patients eventually

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succumb to progressively growing tumors. Further, Lee et al (Journal of Immunology, 1999, vol. 163, pp. 6292-6300) corroborate these findings, stating that although peptide based vaccines can effectively generate a T-cell specific response in the PBMC of cancer patients, said response is not associated with a clinically evidence regression of metastatic melanoma (page 6297, under the heading of "enhancement of vaccine-specific T cell..."). These references serve to demonstrate that induction of a CTL by means of the administered antigens of the invention or the demonstration that said CTL can lyse target cells expressing a tumor associated-antigen in vitro does not constitute evidence that T-lymphocytes would be effective at lysing tumor cells in vivo.

It is noted that the types and stages of generic cancers encompassed by the claims would not be expected to initiate or maintain the same growth kinetics. This is of importance with regard to the teachings of Paul (ibid) on tumor cell escape mechanisms which include rapid growth as a means to overwhelm a slower immune response, (Paul, Fundamental Immunology, (text), 1993, page 1163, second column, first sentence under the heading "Factors Limiting Effective Tumor Immunity" and Table 4) and deficient antigen processing by tumor cells. With regard to the antigen processing, it is unclear whether all patients having a tumor associated antigen would have peripheral T-cells which were specific from the disclosed antigen, as the art teaches that the presence of a small number of tumor cells or the presence of a large number of tumor cells gives rise to tolerance (Paul, page 1166, second column, lines 19-23 under the heading "Sneaking Through"). Based on this observation, it is reasonable to conclude that a small number of slow growing tumor cells would elicit tolerance, and a large number of rapidly growing tumor cells would also elicit tolerance in line with the bi-phasic response reported by Paul. Thus, it appears that the interaction of the tumor cells with the host can produce tolerance by means of clonal deletion within the thymus of said host.

Further, claim 37 is drawn to the administration of any of the peptides in Tables V, VI or VII. It is clear from the specification, that not all of the predicted peptides have the ability to activate CTL against BFA4 expressing target cells because the specification teaches the pooling of the peptides and the deconvolution of the pooled peptides into a few individual peptides responsible for the activity in vitro (page 41, first and second paragraphs). Further the art teaches that "putative epitopes" can be predicted using a computer to scan the sequence of the

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gene (antigen) for amino acid sequences that contain a "motif" or a defined pattern of amino acid residues associated with a particular MHC (HLA) allele, but that upon testing in standard functional assays, the vast majority of these "predicted" epitopes failed to be immunogenic (Burch WO 03/084467 Oct 16, 2004). The specification fails to address how to use peptides which were not able to induce a CTL response against BFA4. Further, claims 31-35 are reliant in part on nucleic acids encoding a fragment of SEQ ID NO:25 and 27. It is noted that these SEQ ID NO are nine-mer peptides. The specification provides no guidance for the selection of smaller fragments of SEQ ID NO:25 or 27 sufficient to treat cancer when expressed via an expression vector.

Given the lack of guidance for all the above issues and the unreliability of treating cancer by the induction of a CTL response targeting said cancer, one of skill in the art would be forced into undue experimentation in order to use the claimed products or carry out the claimed methods for the treatment of cancer.

(D) As drawn to a method of preventing cancer.

Claims 31-35 are drawn in part to a method of preventing cancer comprising administration of an expression vector encoding SEQ ID NO:25 or 27 or a fragment thereof

When given the broadest reasonable interpretation, the "prevention of cancer" include the prevention of a primary cancer in a patient who has yet to develop cancer. This would include the identification of individuals who were going to develop a BFA4 expressing cancer and the administration of the expression vectors of the invention before the development of said cancer. The abstract of Wheeler (Salud p'ublica de M'exico, (1997 Jul-Aug) 39 (4) 283-7) teaches that a cancer vaccine against human papillomavirus for the treatment of cervical cancer requires not only the activation of antigens and overcoming the host response, but the generation of high levels of T and B memory cells; and the persistence of antigens. The instant specification has not provided any teachings regarding the persistence of the tumor antigens in an individual who has yet to develop a specific type of cancer. Further Efferson et al (Anticancer Research, 2005, Vol. 25, pp. 715-24) teach that efficient induction of memory cells is hindered by the lack of information about the relationship between TCR stimulation and the cytokines required for Ag-specific memory CD8+ cells and proliferation and survival. It is noted that the instant specification has not provided any evidence that adequate levels of T and B memory cells would

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persist in an immunized individual who has not developed a cancer, and Efferson et al is clearly discussing a need in the art as of 2005, three years after the priority date of the instant specification, therefore the enablement for how to generate adequate memory T and B cells can not be provided from the general knowledge of in the art. Bachman et al (Journal of Immunology, 2005, Vol. 175, pp. 4677-4685) teach that memory T cells are not a homogeneous population and can be divided into central memory T cells with a substantial capacity for recall proliferation and effector memory T cells with limited recall proliferation capacity. Bachman et al teach that the protective capacity of the different subpopulations of memory T cells vary, and the generation of the subpopulations is influenced by the nature and route of immune challenge. These references serve to demonstrate that the prior art is not mature with respect to how to elicit an effective prophylactic memory cell response that will persist in an individual not harboring a tumor cells and which would function to protect said individual from tumor cell development. Because the specification does not address the issues in the post-filing date art regarding how to elicit an effective memory cell response from the administration of the claimed compositions, and no objective evidence or working examples have been provided, one of skill in the art would be subject to undue experimentation in order to make and use the claimed composition as a vaccine.

(C)As drawn to the NYVAC, ALVAC, ALVAC(2) and TROVAC expression vectors

Claims 4, 5, 9, 10, 14, 15, 19, 20, 24, 25, 29, 30, 34, 35, 47, 48, 52, 53, 57, 58, 62 and 63 require the NYVAC, ALVAC(2), ALVAC and/or TROVAC vector. The specification states that NYVAC, ALVAC and TROVAC were deposited under the terms of the Budapest Treaty (pages 23-24). It is unclear whether one of skill in the art could make the identical vectors of NTVAC, ALVAC and TROVAC without undue experimentation. Therefore the deposit for patent purposes is required in order to satisfy the requirements of 112, first paragraph. However, applicant's referral to the deposits on pages 23-24 of the specification is an insufficient assurance that all required deposits have been made and all the conditions of 37 CFR 1.801-1.809 have been met.

If the deposits are made under the provisions of the Budapest Treaty, filing of an affidavit or declaration by applicant or assignees or a statement by an attorney or record who has

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the authority and control over the conditions of deposit over his/her signature or registration number stating that the deposits have been accepted by an International Depository authority under the provisions of the Budapest Treaty, that all restrictions upon public access to the deposits will be irrevocably removed upon the grant of a patent on this application and that the deposits will be replaced if viable samples cannot be dispensed from the depository as required. This requirement is necessary when deposits are made under the provisions of the Budapest Treaty as the Treaty leaves this specific matter to the discretion of each State.

If the deposits are not made under the provisions of the Budapest Treaty, then in order to certify that the deposits comply with the criteria set forth in 37 CFR 1.801-1.809 regarding availability and permanency of deposits, assurance of compliance is required. Such assurance may be in the form of an affidavit or declaration by applicants or assignees or in the form of a statement by an attorney of record who has the authority and control over the conditions of deposit over his/her signature and registration number averring:

- (a) during the pendency of this application, access to the deposits will be afforded to the Commissioner upon request;
- (b) all restrictions upon the availability to the public of the deposited biological materials will be irrevocably removed upon the granting of a patent on this application;
- (c) the deposits will be maintained in a public depository for a period of at least thirty years from the date of the deposit or for the enforceable life of the patent or for a period of five years after the date of the most recent request for the furnishing of a sample of the deposited biological material, whichever is longest; and
- (d) the deposits will be replaced should they become non-viable or non-replicable.

Amendment of the specification to recite the date of deposit and the complete name and address of the depository is required. As an additional means for completing the record, applicant may submit a copy of the contract with the depository for deposit and maintenance of each deposit.

If deposits are made after the effective filing date of the application for patent in the United States, a verified statement is required from a person in a position to corroborate that the deposited biological materials are producing the NYVAC, ALVAC and TROVAC vectors as described in the specification as filed and are the same as those deposited in the depository,

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stating that the deposited materials are producing identical vectors as NYVAC, ALVAC and TROVAC as described in the specification and were in the applicant's possession at the time the application was filed.

Applicant's attention is directed to *In re: Lundak*, 773 F. 2d.1216, 227 USPQ 90 (CAFC 1985) and 37 CRF 1.801-1.809 for further information concerning deposit practice.

***Claim Rejections - 35 USC § 102***

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(a) the invention was known or used by others in this country, or patented or described in a printed publication in this or a foreign country, before the invention thereof by the applicant for a patent.

(b) the invention was patented or described in a printed publication in this or a foreign country or in public use or on sale in this country, more than one year prior to the date of application for patent in the United States.

(c) the invention was described in a patent granted on an application for patent by another filed in the United States before the invention thereof by the applicant for patent, or on an international application by another who has fulfilled the requirements of paragraphs (1), (2), and (4) of section 371(c) of this title before the invention thereof by the applicant for patent.

(d) the invention was described in (1) an application for patent, published under section 122(b), by another filed in the United States before the invention by the applicant for patent or (2) a patent granted on an application for patent by another filed in the United States before the invention by the applicant for patent, except that an international application filed under the treaty defined in section 351(a) shall have the effects for purposes of this subsection of an application filed in the United States only if the international application designated the United States and was published under Article 21(2) of such treaty in the English language.

The changes made to 35 U.S.C. 102(e) by the American Inventors Protection Act of 1999 (AIPA) and the Intellectual Property and High Technology Technical Amendments Act of 2002 do not apply when the reference is a U.S. patent resulting directly or indirectly from an international application filed before November 29, 2000. Therefore, the prior art date of the reference is determined under 35 U.S.C. 102(e) prior to the amendment by the AIPA (pre-AIPA 35 U.S.C. 102(e)).



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Claims 1-3, 44-46 are rejected under 35 U.S.C. 102(e) as being anticipated by Gish et al (US 6,780,586, reference of the IDS submitted Aug, 16, 2006).

Claim 1 is drawn to an expression vector comprising the nucleic acid sequence of SEQ ID NO:1. Claim 2 embodies the vector of claim 1 which is a viral vector. Claim 3 specifies a retroviral vector. Claim 44 is drawn to an expression vector comprising encoding Q ID NOL2. Claim 45 embodies the vector of claim 44 which is a viral vector. Claim 45 specifies that the vector is a viral vector. Claim 46 embodies the vector of claim 45 wherein the viral vector is a retroviral vector.

Gish et al disclose retroviral vectors comprising SEQ ID NO:1 which encode BFA4 which is identical to the instant SEQ ID NO:2 (column 2, column 14, lines 31-34, column 15, lines 58-61, column 46, example 2)

Claims 1 and 44 are rejected under 35 U.S.C. 102(a) and 35 U.S.C. 102(e) as being anticipated by Birse et al (WO 02/00677).

Birse et al disclose vectors for the recombinant expression of SEQ ID NO:2 encoded by SEQ ID NO:1 (claim 7, page 9, lines 1-2 of [0022], page 1721, first sentence of [0060], page 2456, fifth line from the bottom "190350").

Claims 1-10 are rejected under 35 U.S.C. 102(b) as being anticipated by Tartaglia et al (Vaccine, March 2001, Vol. 19, pp. 2571-2575).

Claim 1 is drawn in part to an expression vector comprising a fragment of SEQ ID NO:1. When given the broadest reasonable interpretation, a fragment can be a single nucleotide of SEQ ID NO:1. Tartaglia et al disclose tumor antigen-ALVAC vectors (page 2572, Table 1) which would comprise a nucleic acid sequence having at least a single nucleotide in common with SEQ ID NO:1. further Tartaglia et al disclose a ALVAC vector expressing both B7.1 and CEA. The nucleic acid encoding B7.1 would have at least a single nucleotide in common with the instant SEQ ID NO:1. The presence of CEA in said ALVAC vector would fulfill the specific requirement of claim 6 requiring a further nucleic acid encoding a tumor antigen.

All claims are rejected.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Karen A. Canella whose telephone number is (571)272-0828. The examiner can normally be reached on 10-6:30 M-F.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Larry Helms can be reached on (571)272-0832. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

Karen A. Canella, Ph.D.

3/17/2006

  
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PRIMARY EXAMINER